

IMP.act

Managing for Microplastics: A Baseline to Inform Policy Stakeholders

Intertidal sampling protocol



May 2019

Table of Contents

Intertidal sampling

PROJECT DETAILS	2
#1 COLLECTION	3
INTERTIDAL SEDIMENT SAMPLING	3
LABORATORY PROCESSING	4
MARINE LITTER SURVEY	5
SAMPLING SITE FORM	6
MATERIAL CHECKLIST	11
#2 APPENDIX (REFERENCES FOR SEDIMENTS AND MICROPLASTICS)	12
#3 REFERENCES	12

Project details

<https://www.joaofrias.com/impact-project>

IMP.act is a project funded by the Irish Research Council under the framework of the Marie Skłodowska-Curie Actions COFUND Collaborative Research Fellowships for a Responsive and Innovative Europe (CAROLINE) scheme (Ref. CLNE/2018/524)

Citation

Frias, J. (2019). Intertidal sampling protocol. <https://doi.org/10.13140/RG.2.2.32410.06082>

#1 Collection

Intertidal sediment sampling

Beaches are dynamic systems with ever-changing conditions and sampling for microplastics should take into consideration that high tide lines can be highly variable over relatively short periods of time. In order to account for this, monitoring surveys should be held, whenever possible, once per season (spring, summer, autumn and winter). To define the sampling area, mark out a 100m transect in width, parallel to the water edge (sea), using a measuring tape or similar and take note of the GPS coordinates on each side of the transect (Figure 1., A and B). This transect will define the sampling area i.e. from the shoreline (low tide, light grey, AC1) to the above the strand line (accumulation zone, dark grey, AC2). In Figure 1 the dark grey line represents the mean height of the spring tide line and the light grey line represents the low tide line. Please note that in many beaches this second tide line might not be always visible on the shore. If the second line does not exist, sample another area outside the accumulation zone.

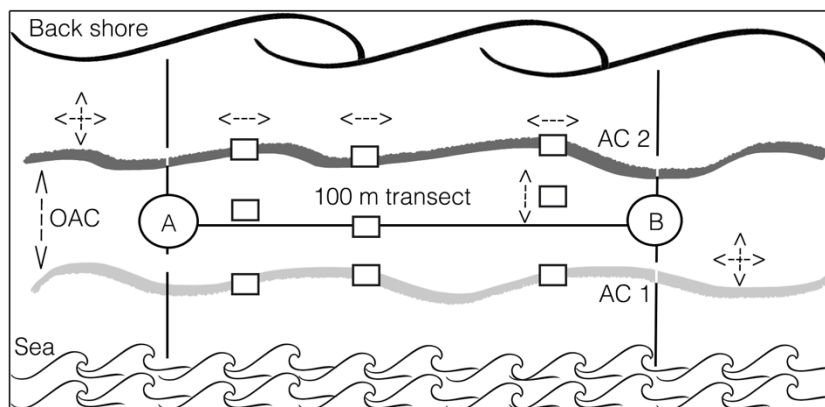


Figure 1 is a visual representative example of what could be drawn in your datasheet. The datasheet in the appendix provides an area for the surveyor to draw a representation of the beach on the day of the survey.

Figure 1 – Example of 100m transect (adapted from OSPAR, 2010 and NOAA, 2013)
(AC – accumulation area, OAC – outside accumulation area)

The sampling design is stratified random so collect a minimum of 3 samples (represented by square areas in Figure 1) along a transect in each high tide line (AC1 and AC2). Make sure you also survey the area between the two high tide lines (OAC).

Mark your sampling unit (30 x 30 cm) using the measuring tape or a quadrat and record the GPS coordinates of each unit. Collect the top 5 cm (total volume of approximately 4,500 cm³ = 4.5 L) of sediment using a small metal shovel and ideally store the sample in labelled glass jars, previously decontaminated. Alternatively, they can be stored in labelled grip- or zip-lock bags. The label should include site name, date, time (if needed), and the code of the sample (AC1a, AC1b, AC1c, OAC1a, OAC1b, etc.). This method will allow to estimate concentration of microplastics both

horizontally and vertically, allowing collected data to be compared with a wide range of studies. Datasheets for this task can be found in the appendix (Form 1).

Laboratory processing

For citizen science purposes, make a sea salt (sodium chloride) saturated solution, as a density separation method. Seawater has an approximately salt concentration of 35 g L⁻¹ and the recommended concentration for this laboratory method is to use a 100 g L⁻¹ concentration. Introduce the sediment sample in a 1L glass beaker, add the saturated seawater solution and stir the solution for approximately 1 minute. Cover the solution with tinfoil to avoid airborne particles to go inside the solution. Wait approximately 20 mins to allow particles to deposit in the bottom. Some plastic polymers will have the tendency to float at the surface (see table A2 in the appendix).

Use a Buchner filtration system (figure 2) and appropriate filters (figure 3) to filter the solution. When dried, observe the filters under a microscope. Use the appropriate form below to register suspicious particles. Register if possible, type, shape, colour, number.



Figure 2 – Buchner filtration system with vacuum pump (credits: Merck Millipore)



Figure 3 – GF/C filters (credits: Fischer Scientific)

Marine litter survey

Once you have collected all the sediment samples from each quadrat and depending on your time availability or litter accumulation of the beach, you can conduct a marine litter survey to quantify and account of larger pieces of marine litter. One of the easiest measures that you can use in a citizen science project or if you are organising a voluntary beach clean-up is to consider collecting everything larger than a bottle cap (approximately 2.5 cm in diameter). In small groups of people (between 3 and 5), one person should be in charge of registering all items into the form. The easiest way to save time is for people to shout out what they are collecting into a large plastic bag. This way, all items will be quantified, and, in the end, you just need to weight the bags. For larger groups of people divide into smaller teams, have 1 person registering and 3 to 4 people collection. The person that is registering should do this tasks for the duration.

The most common marine litter categories can be seen in the specific form below in this document. For more details use the OSPAR report in the reference section, where these methods were adapted from.



Forms

Sampling site form

Country _____ Location area _____

Sampling site code _____ Beach name _____

Date: _____ / _____ / _____ (dd/mm/yyyy) Sampling season: _____

Start time: _____ : _____ AM | PM End time: _____ : _____ AM | PM

Beach Characteristics

Slope: _____ (degrees) Beach length: _____ (m)

Beach substrate: Sand Gravel Boulder. Other _____

Dune Substrate: Sand Gravel Boulder. Other _____

Atmospheric conditions: Strong wind Rain Waves (strong, moderate, low): _____

Did any of the following atmospheric conditions affect OR prevent the sampling on this day?

Storm | hurricane Ice | Frost Fog | Smog Dust- or sand-storm Waves exceptionally high

GPS coordinates:

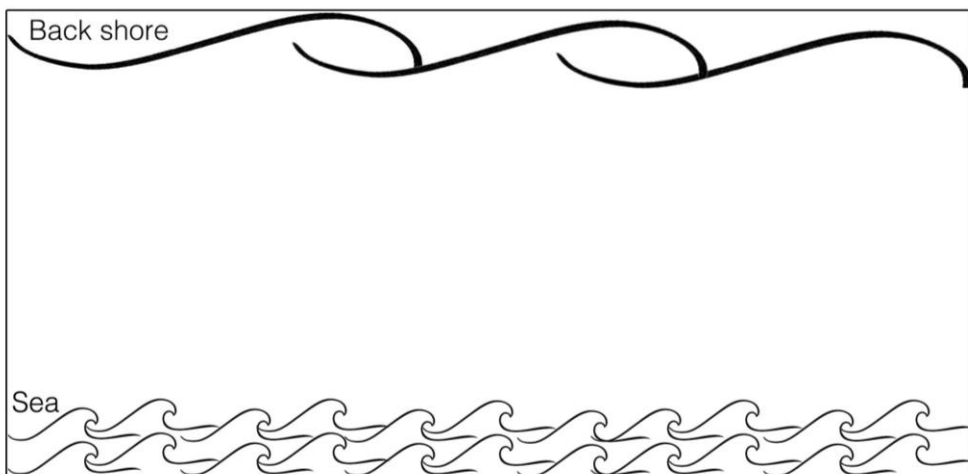
A _____ B _____

AC1 1. _____ 2. _____ 3. _____

AC2 1. _____ 2. _____ 3. _____

OAC 1. _____ 2. _____ 3. _____

SAMPLING (according to scheme in page 3)



1. Draw the high tide lines representing the main accumulation areas (AC1 and AC2);

2. Mark starting point A and finish point B. These should have 100m of distance between them;

3. Draw the squares where sampling was conducted. (For example see Fig. 1).

Comments/Notes: _____

Beach use: Urban Rural Other: _____

Proximity to | Presence of Industry Tourist Attractions (e.g. surf schools) Rivers
 Sewage inputs Harbours/Ports Fishing facilities Marina Other: _____

Factors that might influence the presence of marine anthropogenic litter:

Beach Clean-up activity: Event locally organised by municipality civic movement/NGO.

How frequently is this beach cleaned? Annually Quarterly Bimonthly
 Monthly Weekly Daily

Recent storms or extreme events. Which? _____

Festivals or other events. Which? _____

Impacts on marine fauna

Did you find dead animals? No Yes. How many: _____

Which species?

#1 _____ #3 _____

#2 _____ #4 _____

Were the dead animals entangled in marine litter? No Yes. How many: _____

Please provide more details on the entanglement:

#1 _____ #3 _____

#2 _____ #4 _____

Name of surveyor _____

Contact (e-mail): _____@_____

Was marine litter collected in this activity? Yes No

Notes: _____

Name of surveyor _____

Contact (e-mail): _____@_____

Date: ____/____/20__

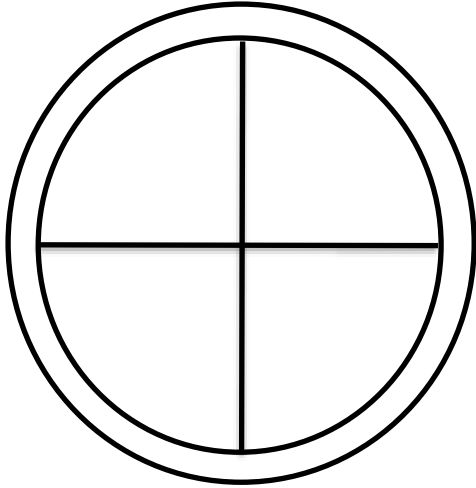
Filter observation form

Sample code _____

Filter no. _____

Date of collection: ____/____/20__

Magnification _____ x



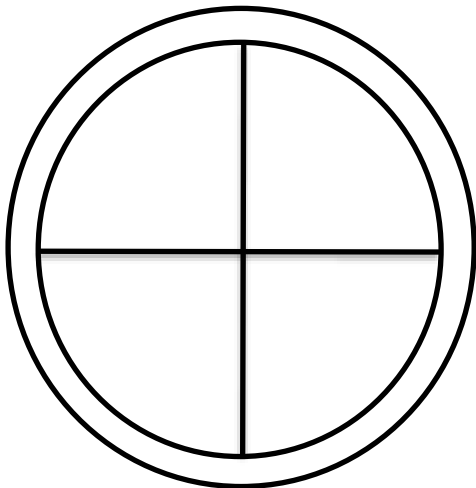
Notes: _____

Sample code _____

Filter no. _____

Date of collection: ____/____/20__

Magnification _____ x

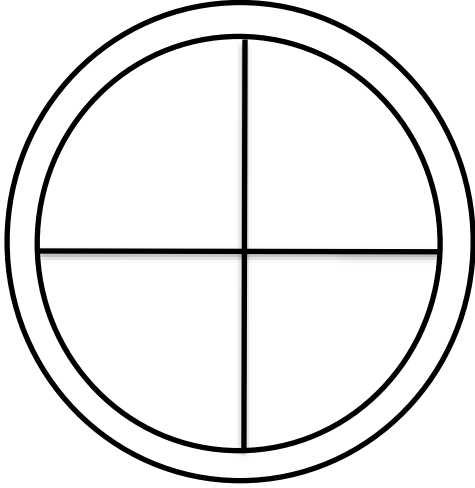


Notes: _____

Date: ____/____/20__

Sample code _____ Filter no. _____

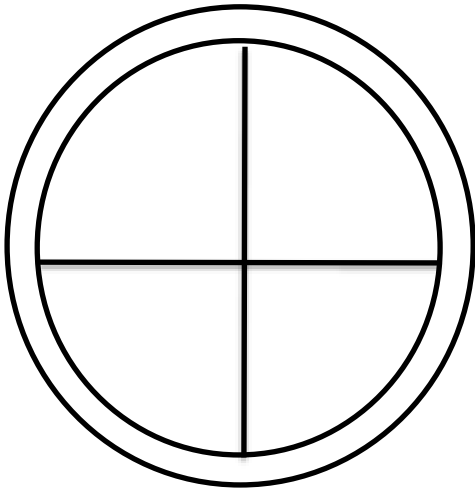
Date of collection: ____/____/20__ Magnification _____ x



Notes: _____

Sample code _____ Filter no. _____

Date of collection: ____/____/20__ Magnification _____ x



Notes: _____

Marine litter collection form

Note: The easiest way to mark the number of items is III ~~IIII~~ = 8 | Use the back of this form if more space is needed

Items most likely to be found:

Cigarette filters: _____	=	<input style="width: 100%;" type="text"/>
Cigarette lighters: _____	=	<input style="width: 100%;" type="text"/>
Beverage bottles (plastic): _____	=	<input style="width: 100%;" type="text"/>
Beverage bottles (glass): _____	=	<input style="width: 100%;" type="text"/>
Beverage cans (metal): _____	=	<input style="width: 100%;" type="text"/>
Bottle caps (plastic): _____	=	<input style="width: 100%;" type="text"/>
Bottle caps (metal): _____	=	<input style="width: 100%;" type="text"/>
Bags (plastic): _____	=	<input style="width: 100%;" type="text"/>
Bags (paper): _____	=	<input style="width: 100%;" type="text"/>
Straws (plastic): _____	=	<input style="width: 100%;" type="text"/>
Cutlery (plastic): _____	=	<input style="width: 100%;" type="text"/>
Plates and cups (plastic): _____	=	<input style="width: 100%;" type="text"/>
Plates and cups (paper): _____	=	<input style="width: 100%;" type="text"/>
Bottle caps Bottle caps (metal): _____	=	<input style="width: 100%;" type="text"/>
Fishing buoys, pots and traps: _____	=	<input style="width: 100%;" type="text"/>
Fishing nets and pieces: _____	=	<input style="width: 100%;" type="text"/>
Fishing line and rope: _____	=	<input style="width: 100%;" type="text"/>
Balloons: _____	=	<input style="width: 100%;" type="text"/>
Condoms: _____	=	<input style="width: 100%;" type="text"/>
Hygiene pads: _____	=	<input style="width: 100%;" type="text"/>
Tampons/tampon applicators: _____	=	<input style="width: 100%;" type="text"/>
Medical tablets: _____	=	<input style="width: 100%;" type="text"/>
Hygienic wipes: _____	=	<input style="width: 100%;" type="text"/>
Syringes: _____	=	<input style="width: 100%;" type="text"/>
Tabaco wraps: _____	=	<input style="width: 100%;" type="text"/>
Food wraps: _____	=	<input style="width: 100%;" type="text"/>
Construction materials (including processed wood): _____	=	<input style="width: 100%;" type="text"/>

Items of local concern:

1. _____	=	<input style="width: 100%;" type="text"/>
2. _____	=	<input style="width: 100%;" type="text"/>
3. _____	=	<input style="width: 100%;" type="text"/>
4. _____	=	<input style="width: 100%;" type="text"/>
5. _____	=	<input style="width: 100%;" type="text"/>

Metrics

Did you register the weight? No Yes.

If so, how much: _____ ? kg lb

Distance cleaned _____ ? km miles

How many people participated? _____

Material Checklist

- Sampling box
- Measuring tape
- Pencils
- Datasheets
- Labels
- Waterproof clipboards
- Photo camera / mobile phone
- GPS / mobile phone
- Metal shovel
- Zip-lock bags
- Permanent markers
- Personal safety equipment (Wellies and wet gear)
- Big plastic bags (25, 50 or 100L depending on the amount of beached litter)
- Gloves (gardening gloves are the recommended due to the thickness)
- Weighing scale
- Transect sticks (3)
- Container for sharp objects
- metal quadrats (3)

#2 Appendix (references for sediments and microplastics)

Table A1 - Densities of common polymers (adapted from Enders *et al.*, 2015)
Density limit using: ■ Sodium chloride and ■ Sodium tungstate dihydrate and all above 1.40 g cm⁻³

Abbreviation	Polymer	CAS no.	Density (g cm ⁻³)
PS	Polystyrene	9003-53-6	0.01 – 1.06
PP	Polypropylene	9003-07-0	0.85 – 0.92
LDPE	Low-density polyethylene	9002-88-4	0.89 – 0.93
EVA	Ethylene Vinyl Acetate	24937-78-8	0.94 – 0.95
HPDE	High-density polyethylene	9002-88-4	0.94 – 0.98
PA	Polyamide	63428-84-2	1.12 – 1.15
PA 6,6	Nylon 6,6	32131-17-2	1.13 – 1.15
PMMA	Poly methyl methacrylate	9011-14-7	1.16 – 1.20
PC	Polycarbonate	25037-45-0	1.20 – 1.22
PU	Polyurethane	9009-54-5	1.20 – 1.26
PET	Polyethylene terephthalate	25038-59-9	1.38 – 1.41
PVC	Polyvinyl chloride	9002-86-2	1.38 – 1.41
PTFE	Polytetrafluoroethylene	9002-84-0	2.10 – 2.30

Polymers until the marked lines are retained by the solutions. Please note that this is a theoretical model and some polymers with higher densities could potentially be found in sediments even using a solution with density lower to 1.40 g cm⁻³.

Table A2 – Buoyancy of common polymers (adapted from Cranford and Quinn, 2017)

Abbreviation	Polymer	Density (g cm ⁻³)	Buoyancy
PS	Polystyrene	0.01 – 1.06	Positive (↑)
PP	Polypropylene	0.85 – 0.92	Positive (↑)
LDPE	Low-density polyethylene	0.89 – 0.93	Positive (↑)
HPDE	High-density polyethylene	0.94 – 0.98	Positive (↑)
Seawater		1.025	
PA	Polyamide	1.12 – 1.15	Negative (↓)
PA 6,6	Nylon 6,6	1.13 – 1.15	Negative (↓)
PMMA	Poly methyl methacrylate	1.16 – 1.20	Negative (↓)
PC	Polycarbonate	1.20 – 1.22	Negative (↓)
PU	Polyurethane	1.20 – 1.26	Negative (↓)
PET	Polyethylene terephthalate	1.38 – 1.41	Negative (↓)
PVC	Polyvinyl chloride	1.38 – 1.41	Negative (↓)
PTFE	Polytetrafluoroethylene	2.10 – 2.30	Negative (↓)

Polymer density might vary with additives added during production, and therefore this table is a theoretical model.

Tables available at <https://www.researchgate.net/publication/326552185>

#3 References

1. OSPAR (2010). Guideline for Monitoring Marine Litter on the Beaches in the OSPAR Maritime Area. ISBN: 90 3631 973 9. Available @ <https://www.ospar.org/documents?v=7260>
2. Frias *et al.*, (2018). Standardised protocol for monitoring microplastics in sediments. JPI-Oceans, published in ResearchGate. Available @ <https://doi.org/10.13140/RG.2.2.36256.89601/1>



Managing for Microplastics: A Baseline to Inform Policy Stakeholders

This project contributes to the following
United Nations Sustainable Development Goals



May 2019